I. INTRODUCTION

Diarrhoea and gastro-enteritis in human beings can be caused by viruses (Rotavirus, Adenovirus, Astrovirus, Norwalk virus, etc), bacteria such as Salmonella and E. coli, and protozoa such as Cryptosporidium and Giardia. Viruses cause 45% of the diarrhoea in children under 1 year of age and 40% of the diarrhoea in children under 4. The prevalence of Adenovirus is 4-12%. This makes it the second leading cause of viral enteritis in children under two years of age.

Infection occurs via the faecal-oral route, but can result from the inhalation of aerosols as well. The incubation period lasts from 5 to 8 days and the symptoms of the stomach and intestinal inflammation are watery diarrhoea, vomiting, fever, and abdominal cramps.

The Adenoviruses are divided into six subgroups labelled A to F. Subgroup F is the most frequently involved in paediatric gastro-enteritis.

II. PRINCIPLE OF THE TEST

This test is ready to use and is based on the homogeneous membrane system technology with colloidal gold particles. The faecal specimen must be diluted in the dilution buffer that is supplied with the test. A nitrocellulose membrane is sensitized with antibody to Adenovirus.

The test’s specificity is ensured by a monoclonal antibody specific to the Hexon antigens of Adenovirus A through F that is conjugated to the colloidal gold. This conjugate is insolubilized on a polyester membrane.

When the strip is dipped into the liquid phase of the faecal suspension, the resolubilized conjugate migrates with the sample by passive diffusion and the conjugate and sample material come into contact with the anti-Adenovirus monoclonal antibody adsorbed to the nitrocellulose. If the sample contains Adenovirus, the conjugate-Adenovirus complex remains bound to the anti-Adenovirus monoclonal antibody. The result – in the form of a red line that forms on the strip – is visible within 10 minutes. The solution continues to migrate to encounter a control reagent that binds a control conjugate, thereby producing a second red line and confirming that the test is working properly.

III. REAGENTS AND MATERIALS

Each kit contains Adeno-strips, dilution buffer and optional components (for C-1502):

1. Adeno-Strip strips
   Each strip is sensitized with a mouse monoclonal anti-Adenovirus antibody and with a control reagent. The purified reagents are adsorbed to the nitrocellulose.
   The anti-Adenovirus conjugate is produced with mouse monoclonal antibody directed against the Hexon antigens of groups A through F. This purified antibody is conjugated to colloidal gold particles.
   These strips come in a bottle or a pouch with a desiccant.

2. Dilution buffer (15 ml)
   Saline solution buffered to pH 7.5 with Tris and containing EDTA, NaN3 (<0.1%), a detergent, and charged proteins.

3. Instruction for use (1)

4. Required materials (supplied with C-1502):
   - 3 or 5 ml test tubes;
   - inoculating loops for taking the faecal samples.
   - cardboard rack

Materials to be ordered separately
- Adenovirus Control Test (Ref.: C-1082)

IV. SPECIAL PRECAUTIONS

- All operations linked to the use of the test must be performed in accordance with Good Laboratory Practices (GLP).
- The Adeno-Strips are for in vitro diagnostic only.
- Avoid touching the nitrocellulose with your fingers.
- Wear gloves when handling the samples.
- Dispose of gloves, swabs, test tubes, and sensitized strips in accordance with GLP.
- Never use reagents from another kit.
- If strips are stored in container, the container must be recapped resealed as soon as the necessary number of strips for the operation has been removed, since the strips are sensitive to humidity. Make sure that the desiccant is present.
  - If strips are stored in individual pouches, pouch must be opened with care to avoid damaging the strip.
- Two green lines indicate the antibody adsorption sites. They disappear in the course of the test.
Discard the buffer solution if it is contaminated with bacteria or mould.

To avoid diluting the colloidal gold conjugate in the solution, take care not to immerse the strip above the line placed under the green arrow.

V. STORAGE
An unopened Adeno-Strip kit may be kept at between 4 and 37°C and used until the shelf-life date on the packaging. The strips remain stable for 15 weeks (in the closed container) after the bottle is opened if they are kept at between 4 and 37°C and in a dry environment. The Adeno-Strips and the buffer must not be frozen.

VI. SAMPLES
The stool samples must be tested as soon after they are collected as possible. If necessary, they may be stored at 2-8°C for 1 week or -20°C for longer periods of time. Make sure that the samples are not treated with solutions containing formaldehyde or its derivatives.

VII. PROCEDURE
Preparations:
If the Adeno-Strip kit was kept at 4°C, let all the reagents warm up to room temperature before proceeding with the test. Write the patient’s name or specimen number on the test tube (foresee one test tube per sample). Place the marked test tubes in a rack.

Procedure:
1. Add 0.5 ml or 15 drops of the dilution buffer solution to each tube.
2. Dip the inoculating loop containing the stool sample into the tube. The dilution ratio must be at most 4% w/v. For liquid samples, take 2 loops of 10 µL; for solid samples, take 1 loop.
3. Stir to homogenize the solution and let stand for 1-2 minutes.
4. Discard the inoculating loop and immerse the sensitized strip in the direction indicated by the green arrow.
5. Let react for 10 minutes. Results must be read on wet strips after 10 minutes incubation.

VIII. INTERPRETING THE RESULTS
The results are to be interpreted as follows:

1 upper line = negative
2 lines = positive
0 line = invalid*

* The absence of the control line, which is the upper line, makes the result invalid. In this case, the sample must be retested.

The intensity of the test line may vary according to the quantity of antigens found in the sample. Any signal, even weak, on the test line must be regarded as a positive result. Nevertheless, the test is qualitative and cannot predict the quantity of antigens present in the sample. The clinical presentation and other test results must be taken into consideration to establish diagnosis. During the drying process, a very faint shadow may appear at the test line. It should not be regarded as a positive result.

To store the results, let the strip dry after removing the absorbent material at its base.

IX. QUALITY CONTROL
In accordance with Good Laboratory Practices, we recommend checking the test’s performance regularly in line with the laboratory’s requirements. To do this, the Adenovirus Control Test (C-1082), in which the strip is immersed, may be used. Refer to the C-1082 package insert.

X. PERFORMANCES
A. Detection limit:
For Adenovirus, the analytical sensitivity has been realised with a purified adenovirus-5 antigen and has been evaluated at about 3.9 x 10^8 vp/mL.

B. Sensitivity - Specificity (Correlation):
The kit was validated at the Mons-Hainaut Hygiene Institute (Belgium) by comparing the Adeno-Strip’s results with those of an ELISA test. The Adeno-Strip kit’s sensitivity and specificity were tested on 432 stool samples. The following results were obtained:
## ELISA Adeno-Strip

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<th>Positive</th>
<th>Negative</th>
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<td>25</td>
<td>2</td>
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<tr>
<td>Negative</td>
<td>2</td>
<td>403</td>
<td>405</td>
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<tr>
<td><strong>Total</strong></td>
<td>27</td>
<td>405</td>
<td>432</td>
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- **Sensitivity** = 92.6% (25/27)
- **Specificity** = 99.5% (403/405)
- **Accuracy (Concordance)** = 99.1% (428/432)

(N = 432)

### C. Accuracy:

To check the intra-lot accuracy, same positive sample and a buffer solution have been processed 15 times on sticks of the same production lot in the same experimental conditions. All observed results were correct as expected.

To check the inter-lot accuracy, some samples (positive and buffer) were processed on five different production lots. All results were correct as expected.

### D. Interference:

Cross-reactivity to samples positive for the following pathogens was tested and found to be negative:

- Cryptosporidium parvum (n = 9)
- Campylobacter jejuni (n = 10)
- Giardia lamblia (n = 10)
- Rotavirus (n = 25)
- E. coli 0157: H7 (n = 2)
- Salmonella typhimurium (n = 1)
- Samonella enteritidis (n = 1)
- Yersinia enterocolitica (n = 3)
- Helicobacter pylori (n = 1)
- Aeromonas hydrophila (n = 1)

### XI. LIMITS OF THE KIT

Adeno-Strip kit results must be compared with all other available clinical and laboratory information.

A positive test does not rule out the possibility that other pathogens may be present.

The Adeno-Strip is an acute-phase screening test. Stool specimens that are collected after this phase may contain antigen titres below the reagent’s sensitivity threshold.

### XII. TECHNICAL PROBLEMS / COMPLAINTS

If you encounter a technical problem, or if performances do not correspond with those indicated in this package insert:

1. Record the lot No of the kit in question
2. If necessary, store the problematic sample in the freezer as soon as possible
3. Contact Coris BioConcept or your local distributor

### XIII. BIBLIOGRAPHIC REFERENCES

1. Improvement of the specificity of enzyme immunoassays for the detection of Rotavirus and Adenovirus in fecal specimens.
   Rabeneau, H., Knoll,B., Allwin,R., Doerr,H.W. And Weber, B.
   Intervirology, 1998 ; 41(2-3) : 55-62
2. Comparison of detection methods for Adenovirus from enteric clinical specimens.
   Ahluwalia,GS., Scott-Taylor,TH., Klosko,B. and Hammond, GW.
3. Evaluation of rapid culture centrifugation method for Adenovirus detection in stools.
   Durepaire,N., Ranger-Roger,S. and Denis,F.
4. Importance of Rotavirus and Adenovirus types 40 and 41 in acute gastroenteritidis in Korean children.
   Kim,KH., Yang,JM., Joo,SL., Cho,YG., Glass,Rt. And Cho,YJ.
5. Gastroenteritidis caused by Adenoviruses 40/41 : epidemiological and clinical aspects.
   Pena,MJ., Elcuaz, R., Suarez,J. and Lafarga, B.
6. Prevalence of group A Rotavirus, human calicivirus, astrovirus and adenovirus type 40 and 41 infections among children with acute gastroenteritis in Dijon, France.
   Ben,F., Facsia,P., Dauvergne,M., Tenebaum,D., Planson,H., Petion,AM., Pothier,P. and Kohli,E.

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<tr>
<td></td>
<td>Contains sufficient for &lt;n&gt; tests</td>
<td>DIL SPE Diluent specimen</td>
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<tr>
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<td>Keep intact, dry</td>
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<tr>
<td>DIL AS</td>
<td>Diluent assay</td>
<td>CONT NaN3 Contains Sodium azide</td>
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![Diagram](image-url)